

Product Information Version 2.0

ZEISS Celldiscoverer 7Your Automated Platform for Live Cell Imaging



Your Automated Platform for Live Cell Imaging

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Often in life sciences research, the data you are after will only be revealed using multiple runs of experiments and complex equipment. Automation can be the only way to get there. With Celldiscoverer 7, you can combine the easy-to-use automation of a boxed microscope with the image quality and flexibility of a classic inverted research microscope. Celldiscoverer 7 calibrates itself, then detects and focuses on your samples while the optics adjust themselves. Leaving you free to get on with other projects. Whether working with 2D or 3D cell cultures, tissue sections or small model organisms, you will acquire better data in shorter times with this reliable automated research platform. What's more, you can enhance your Celldiscoverer 7 with optical sectioning to get more information from your three-dimensional samples. It's your choice whether you opt for confocal imaging with LSM 900 and Airyscan 2 or fast GPU deconvolution.





Simpler. More Intelligent. More Integrated.

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A Flexible Platform

Celldiscoverer 7 is a fully integrated high-end imaging system. It comes with various incubation and detection options so you can tailor the system to your applications. Go for fast, sensitive sCMOS or EMCCD cameras when performing your most demanding live cell experiments and rapid time-lapse recordings. To get better data from your 3D samples, simply add the optional LSM 900 with Airyscan 2 for confocal imaging, or fast GPU-based deconvolution. Get all these benefits and more with the in-built flexibility of Celldiscoverer 7.

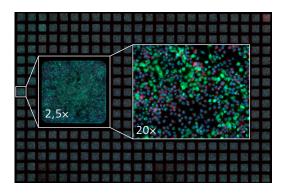
Top Quality Data from Your Samples

For demanding long-term, time-lapse imaging, Celldiscoverer 7 gives you the advantage of Auto-immersion and a hardware-based focus that finds and keeps the focus automatically after detecting the thickness and optical properties of the sample carrier. Autocorr objectives then correct spherical aberrations to deliver crisp contrast and high resolution every time. Get image quality like you've never seen before – no need to adjust manually. Keep your cells happy and they'll deliver unbiased data: Celldiscoverer 7 provides a range of integrated incubation options to create just the right environment. The improved optical design resolves more details in large fields of view.

Reproducible Results Made Easy

As soon as you start imaging, automatic calibration routines take over to ensure reproducible results. Check the current status and follow progress of your experiments on the touchscreen. With barcode recognition you can identify your sample, sample carrier and even the type of experiment. If you don't work with barcodes, an automatic preview scan will identify the sample carrier and calibrate it. ZEISS predictive service offers lasting and optimal instrument performance for increased system uptime and reliable results.







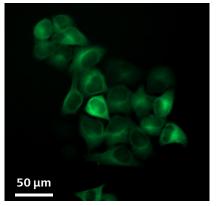
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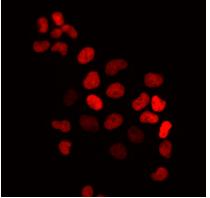
An Easy-to-Use Integrated Microscope

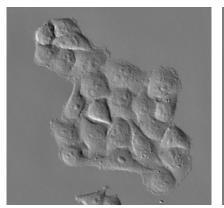
Observing live samples over a number of days or imaging lots of multiwell plates really puts your microscope through its paces. To get reproducible, unbiased data, you must control environmental conditions such as light, temperature, CO₂ etc. That's why Celldiscoverer 7 brings you a unique combination of a stable box, darkroom and integrated inverted research microscope with optional incubation. It simplifies your laboratory setup and makes work more comfortable.

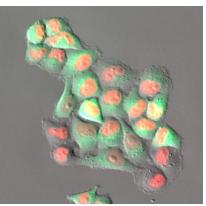
All Celldiscoverer 7 components are optimized for hassle-free automated imaging. New users and multi-user facilities especially will enjoy the in-built automation and usability features when setting up complex experiments. You'll systematically avoid accidental hardware changes that might lead to biased data or even damage your microscope. And Celldiscoverer 7 can make you more productive, too: expect better data in shorter times, with less training and maintenance. What's more, as your needs grow you can expand Celldiscoverer 7 with confocal technology, external cameras, deconvolution, additional environmental control – whatever you need for the challenge of live cell observation.









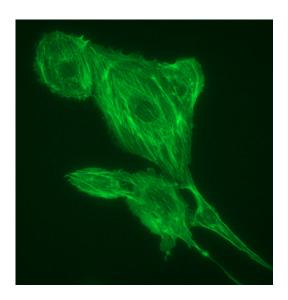


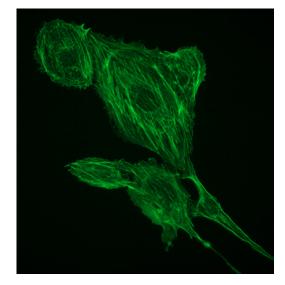
72 h cell growth assay using a waterimmersion objective. HeLa Kyoto cells expressing H2B-mCherry Tubulin eGFP (Neumann et al., Nature 2010 Apr.1.; 464(7289):721-7) imaged every 15 minutes for 72 hours using Autoimmersion; individual channels of the green (eGFP) and red (mCherry) fluorescence and the phase-gradient-contrast as well as an overlay. Sample courtesy of I. Charapitsa, Chemical Biology Core Facility, EMBL, Heidelberg, Germany

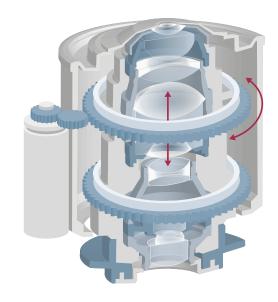
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ZEISS Celldiscoverer 7 Recognizes and Adapts Automatically to Your Samples

Live cell imaging requires objectives with high numerical apertures. Those objectives will only deliver high contrast and sensitivity if their optics can adapt to variations in bottom thickness or to the material of different sample carriers. With Celldiscoverer 7 you're now free to use Petri dishes, chamber slides, multiwell plates, plastic or glass, thin or thick vessel bottoms, low skirt or high skirt plates. Automatic sample recognition detects all relevant vessel features while loading your sample. Then Autocorr adjusts the correction ring of the objective to compensate for spherical aberrations. Find Focus automatically places your sample in focus and Definite Focus keeps it there. It's never been easier to get crisp images with low phototoxicity from deep inside your sample.







Left image shows spherical aberration due to unadjusted optics. Right image shows the same structure using an Autocorr objective. The correction results in increased contrast, resolution and intensity, providing low phototoxicity. The images show tubulin in FluoCell prepared slide #1. Sample courtesy of Invitrogen, Thermo Fisher Scientific Inc.

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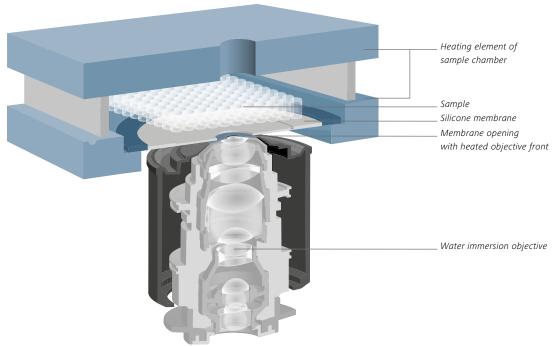
There Is No Life Without Water ...

... and no live cell imaging without water immersion. In life sciences, cell biology or screening applications, your samples mostly consist of water and/or will be mounted in aqueous solutions. Celldiscoverer 7 combines an outstanding water immersion objective with rapid automated immersion supply and removal.

A unique elastic silicon membrane fits perfectly between the objective and sample chamber. The silicon membrane simultaneously seals the sample chamber to avoid unnecessary airflow while protecting the system from potential liquid spillage. Just select the water immersion objective and water is supplied instantly to the front lens. Within seconds the immersion is building up and the lens is ready to use. When you switch back to one of Celldiscoverer 7's dry objectives, the immersion water is automatically removed. Until now, automated imaging systems often struggled as the immersion water quickly evaporated. Celldiscoverer 7 solves that problem by automatically monitoring the immersion and adding water in regular intervals, as needed. With Celldiscoverer 7 you can perform unbiased live cell experiments at 37 °C over several days or carry out extensive scanning processes on multiwell plates.

By adapting the refractive index of your imaging system to the samples, you'll achieve more efficient light collection and increased sensitivity. And less phototoxicity significantly increases viability of even your most challenging living samples.





A silicone membrane allows automatic water immersion and seals the sample chamber.

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Get in Focus, Then Stay in Focus

Use the hardware-based Find Focus function to automatically focus your sample and find your region of interest quickly with just a single click. This significantly reduces the time to your first image and minimizes sample illumination.

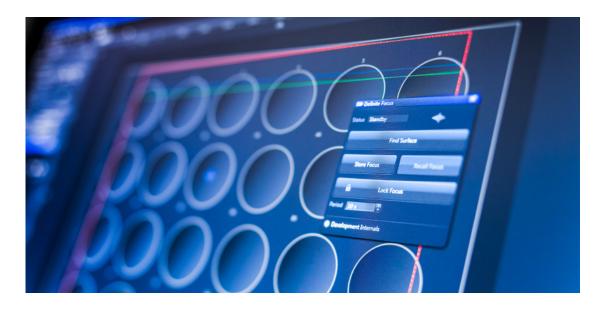
Then select Definite Focus to maintain the focal position throughout your experiments, whether it takes a few seconds or several days.

Or combine both methods with the powerful content-based autofocus of ZEN imaging software. Celldiscoverer 7 can automatically create focus maps for multiple positions in long-term time-lapse experiments. Simply choose the best focus strategy for the experiment at hand.

Move to the Edge ...

... but not one step more, thanks to the Adaptive Lens Guard. High optical performance often compromises on the possible scanning area. Celldiscoverer 7 with its Adaptive Lens Guard protects the objective from collisions with your sample vessel or hardware components, automatically maximizing the available scanning area. Bottom thickness, skirt height and lateral dimensions are important geometrical features of the different sample carrier types – especially when

working with multiwell plates. Celldiscoverer 7 automatically detects these features and adapts accordingly. It also calculates the maximal possible scanning area automatically, depending on the individual sample carrier, objective and current focus position in your experiment. The available scanning area is always indicated on your monitor. Change your experimental parameters and the scanning area will adapt automatically, in real time.

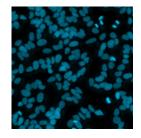


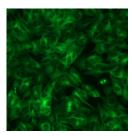
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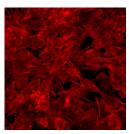
Capitalize on LED-Technology for Live Cell Imaging

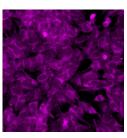
Celldiscoverer 7 brings you all the advantages of LED-technology for efficient illumination with low phototoxicity, fast switching times and long-term stability. That's what delivers gentle imaging, increased throughput and reproducible results. The fluorescence excitation unit combines up to seven LEDs for maximum flexibility in the choice of dyes – from deep blue to far red. All LEDs are hardware-triggered for precise, fast illumination. During sample navigation LEDs are tightly synchronized with camera frame rates. An automated rectangular excitation field stop illuminates only the active field of view, greatly reducing phototoxicity and fluorescence bleaching. Use highefficiency multi-bandpass filter sets for fast acquisition of multiple fluorescent channels. Celldiscoverer 7 simply switches LEDs on/off – without moving any mechanical parts – so you get high-speed multi-channel imaging, even when combined with transmitted light.





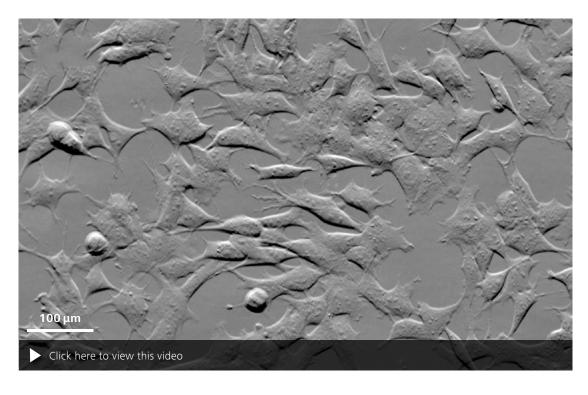






SH-SY5Y cells cultured on a 384 microwell plate. Multichannel image at a single position using the 20×/0.95 objective. Extended depth of focus from Z-stack. Hoechst – Chromatin (blue), anti-alpha-tubulin antibody FITC for alphas tubulin (green), Phalloidine for actin (red), MitoTracker Deep Red for mitochondria (purple). Sample courtesy of P. Denner, Core Research Facilities, German Center of Neurodegenerative Diseases (DZNE), Bonn, Germany.

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SH-SY5Y cells cultured on a 384 microwell plate. Timelapse has been acquired using 20x magnification and phase gradient contrast. Sample and assay courtesy of P. Denner, Core Research Facilities, German Center of Neurodegenerative Diseases (DZNE), Bonn, Germany.

Use a Novel Transmitted Light Contrast

With Celldiscoverer 7 you can use transmitted light brightfield and phase gradient contrast. This novel relief contrast adapts automatically to the sample carrier geometry, providing excellent contrast to the very edge of the vessel. It's fully compatible with all objectives, filter sets and sample carriers. This contrasting method stays robust, even against liquid meniscus or plastic lids. Use the far-red transmitted light LED for gentle

imaging at very high speeds. You can perform applications based on label-free assays or let the system automatically combine transmitted light with multiple fluorescence channels. All multibandpass filter sets support the combination of transmitted light and fluorescence, without reducing sensitivity or speed. On top of that, this unique motorized transmitted light unit allows dispensing directly on the optical axis, without

disturbing the environmental conditions. The dispensing unit is always integrated. As soon as you open the hatch on top of your Celldiscoverer 7, the transmitted light unit will automatically change place with the dispensing unit. You now have direct on-axis access to the specimen for pipetting. You can add agents while maintaining continuous physiological conditions.

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ZEISS Plan-APOCHROMAT 5×/0.35 Objective

This objective is your choice for efficient sample navigation. It creates impressive overview images by delivering an unparalleled information density in a single shot, especially in combination with the microscope camera Axiocam 512 mono. Many screening applications will strongly benefit from the high resolution on large fields. The objective easily handles thin and thick vessel bottom made of glass or plastic. In combination with the built-in magnification changer it combines the benefits of three different objectives into one: $2.5 \times /0.12$, $5 \times /0.25$ and $10 \times /0.35$ — at a fixed working distance.

ZEISS Plan-APOCHROMAT 20×/0.7 Autocorr Objective

From thin to thick, from plastic to glass – this objective adapts automatically to every sample you load on your Celldiscoverer 7. It delivers an unparalleled numerical aperture of 0.7 through 1.2 mm plastic bottom without compromising image resolution and contrast. This tremendous flexibility will make the lens your multipurpose objective, especially if you would like to image cells, which can only grow on plastic bottom. In combination with the built-in magnification changer this objective combines the benefits of three different objectives into one: 10×/0.35, 20×/0.7 and a 40×/0.7 – at a fixed working distance.

ZEISS Plan-APOCHROMAT 20×/0.95 Autocorr Objective

This objective delivers high numerical apertures without applying immersion. It is optimized for thin vessel bottoms. No matter if your cells prefer glass or plastic – this objective will adapt to bottom material and thickness variations. With the increased sensitivity this objective is ideal to generate crisp images on large areas or multiple positions at high speed. In combination with the built-in magnification changer this objective combines the benefits of three different objectives into one: $10\times/0.5$, $20\times/0.8$ and $40\times/0.95$ – at a fixed working distance.

ZEISS Plan-APOCHROMAT 50×/1.2 W Autocorr and Autoimmersion Objective

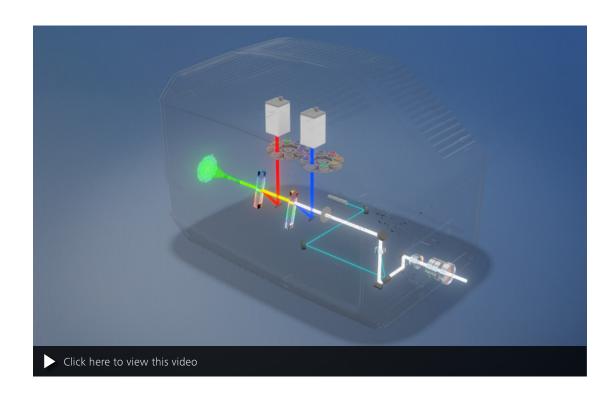
This objective delivers high light collection efficiency and resolution. In combination with the Autoimmersion function it matches perfectly to samples in aqueous solution. Since it reduces phototoxicity to a minimum, it's your choice for your most demanding life cell imaging applications, e.g. long-term imaging of subcellular structures. Optimized for thin bottoms it adapts automatically to the bottom material and thickness. No matter which field of view you prefer, this objective will deliver a constant numerical aperture of 1.2 and combines the benefits of three different objectives into one: 25×/1.2, 50×/1.2 and 100×/1.2 – at a fixed working distance.

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Perform Automated Gentle and Fast Confocal 3D Imaging

Life happens in 3D – and your research often calls for optical sectioning to image your samples with best possible contrast and resolution. Now you can add LSM 900 with Airyscan 2 to your Celldiscoverer 7. You get the best of both worlds: ease of use and automation from a fully integrated microscope platform and the superb confocal image quality and flexibility of the LSM 9 family with Airyscan 2. You perform superresolution 3D imaging with up to 1.5× resolution improvement. And you easily separate multiple labels with spectral imaging. You can now analyze dynamic processes with photomanipulation for FRAP, FRET or related techniques. It's never been easier to precisely connect widefield and confocal images. Fast mixed-mode acquisition simplifies and speeds up your workflow and gives you unique insights into your sample.

The elegant beam path design of your LSM 900 with Airyscan 2 gives you spectral flexibility. Each single component is optimized for highest sensitivity and contrast. Instead of losing light at a closed pinhole, Airyscan 2 collects more emission light and extracts more spatial information than classic confocals. You can use Airyscan 2 with all high NA dry or water immersion objectives.

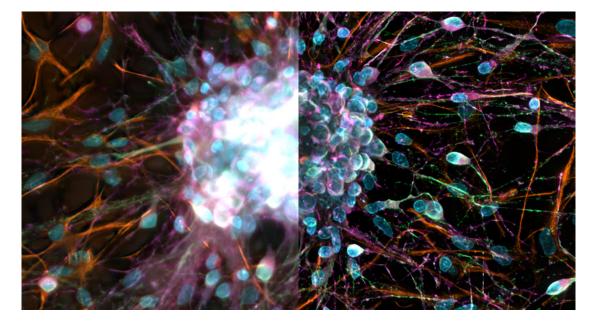


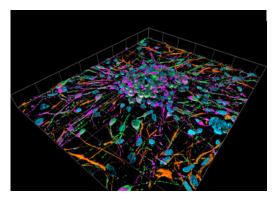
Your live cell imaging benefits from superb image quality with greatly reduced phototoxicity.

Use low magnification widefield to quickly pre-scan your complete sample, then identify regions of interest. It's so easy to then image those regions with LSM 900 and Airyscan 2 for optical sectioning with superresolution. The new Multiplex mode

for Airyscan 2 employs smart detection schemes for this unique area detector. This parallelization allows to image two times faster, while keeping best resolution and SNR. Use this mode to image dynamic processes, or to achieve higher throughput and productivity.

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Rat cortical primary culture. Antibody staining of bIII-tubulin (Cy2, green), Nestin (Cy3, red) and DCX (Cy5, purple), nuclei stained with DAPI (blue). 3D reconstruction of the deconvolved Z-stack (shadow projection). Sample courtesy of H. Braun, LSM Bioanalytik GmbH, Magdeburg, Germany.

Comparison between widefield (left) and deconvolved (right) Z-stack projection using GPU-based Deconvolution.

Get More Details with Deconvolution

When imaging three-dimensional samples, outof-focus light sometimes blurs your structure of interest. For these images, you need deconvolution – a combined optical and mathematical method – to increase contrast and improve the signal-to-noise ratio and resolution. With Celldiscoverer 7 it is easier than ever before to first acquire a Z-stack of your samples and then deconvolve the image to reassign all detected photons to their origin. With ZEN imaging software you use advanced deconvolution algorithms, including a novel approach with depth variant point-spread-functions for deep imaging. Combine this with Celldiscoverer 7's unique Autocorr objectives and you will get excellent results from thicker samples, e.g. 3D-cell culture. And you will get

them up to 30 times faster than with the traditional technology that works on your processing PC's RAM, thanks to Celldiscoverer 7's new GPU-accelerated, parallel CUDA processing. Use the increased speed to extract maximum information from the large datasets you acquired in those demanding long-term, time-lapse or multiwell screening applications.

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Easily achieve stable environmental conditions for your demanding live-cell experiments. You can control the temperature with the optional heating unit or a Julabo cooling circulator. In combination with a humidifier, optional CO₂ and/or O₂ module you control athmospheric conditions.



Depending on your most common imaging needs, you can now choose between Axiocam 506 mono or Axiocam 512 mono.



No matter if you choose a ZEISS Axiocam microscope or a third party camera – if you have to increase acquisition speed and sensitivity for special applications, Celldiscoverer's additional camera port provides the flexibility you need.



Your Celldiscoverer 7 can load multiwell plates, dishes, chamber-slides or standard slides.
All sample holders are optimized for large scanning areas, fully compatible with water immersion and autoclavable.



Celldiscoverer 7 allows you to run perfusion experiments efficiently, while maintaining homogenous and stable environmental conditions.



Celldiscoverer 7 offers an effective way to keep the sample chamber clean. The insert plate for UV disinfection is automatically recognized by the system and you start the disinfection workflow via the touchscreen.

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ZEN Imaging Software Shortens the Path to Your Goal

ZEN – ZEISS Efficient Navigation – is the single user interface you will see on all imaging systems from ZEISS. ZEN imaging software leads you simply and quickly to the result.

At all times you see which options the system is making available to you and which step is

appropriate to take next. ZEN makes it easy to operate every imaging system from ZEISS correctly and intuitively. As a result you save time, reduce training and support costs, and get faster answers to your questions.

With Celldiscoverer 7 you profit from advanced automation features:

- Simple and intuitive carrier-based navigation via mouse and keyboard
- A dedicated automation wizard to create scan profiles for routine or reoccurring tasks
- A range of hardware- and software-based focus strategies to set up even complex multiposition experiments
- Fast overview images. Create an overview of your cells just once, then there's no need to expose them to unnecessary light doses during experiment setup.
- Cell viability put first with samples illuminated only as long as the camera acquires an image
- An optimized CZI file format for large datasets and seamless integration into existing image analysis workflows
- Open interfaces. Use your CZI dataset in all major software packages that use the BioFormats library, e.g., Fiji, Python, Matlab, Icy, Knime, Imaris, Arivis.

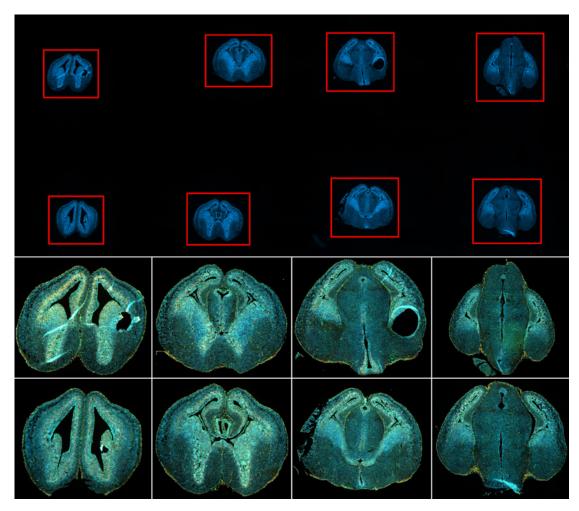
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OAD is Your Interface to ZEN Imaging Software

- Use Python scripts to customize and automate your workflows.
- Integrate external image analysis applications into your workflows.
- Exchange image data with external programs like ImageJ, Fiji, MATLAB, KNIME or Python.
- Use feedback for smart experiments.
- Get more reliable data in less time.
 It's your choice.



OAD enables the analysis of data acquired with ZEN imaging software by other programs like ImageJ. Transfer your results back to ZEN for further analysis and display.



The result of overview scan using low magnification (top panel) was used to automatically detect the brain slices via image analysis. The results (XYZ position and the height/width of detected objects) were used in a automated subsequent scan using a high NA objectives, where the system carried out an individual tile scan for every detected object in a complete automated fashion without any additional user interaction. Sample courtesy of P. Grigaravicius, FLI – Leibniz Institute on Aging, Jena, Germany.

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ZEISS ZEN Connect Lets You Overlay and Organize Images from Any Source.

Connect All Your Multimodal Data to Expand Correlative Microscopy

Expanding classic correlative microscopy, ZEN Connect is open to all your images: you can load complex multidimensional images as easily as simple overview images from your mobile phone. It makes no difference whether your imaging technology is from ZEISS or from third parties. All image data can be aligned, overlayed and shown in context. So long as your external images adhere to the well-established Bio-Formats standard, ZEN Connect will even keep their metadata.

Acquire Overview Images for Easy Navigation

Image your sample with a ZEISS stereo microscope or any other low magnification system.

Then move to your high-resolution system of choice. With ZEN Connect you only need to align it once, then use the overview image to navigate and find your ROIs. All subsequent high-resolution images will be shown in context as you zoom in and out across the borders of resolution domains and imaging technologies. A single click on the overview image brings your stage to the right position to examine or reevaluate any of your ROIs with the full image overlay.

Smart Data Management

All the images you acquire with ZEN Connect are saved in well-structured database projects, complete with an intuitive label attached automatically to each image file. You'll always stay on top of things – during your experiments as well as months afterwards when analyzing your work. It's easy to find all your overlay images and their connected datasets. You can even search for microscope type and imaging parameters with the new filter function of ZEN Connect.



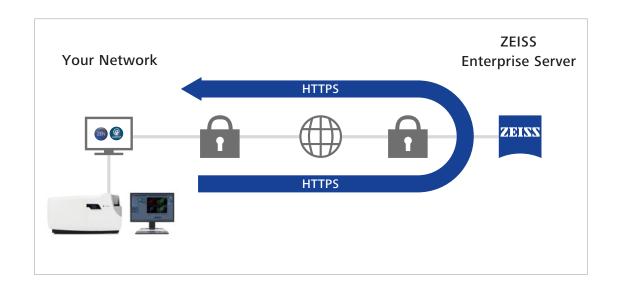
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ZEISS Predictive Service Maximizes System Uptime

Once connected to your network and activated, this advanced technology will automatically track the health status of your instrument and collect system log files in the background to improve remote diagnosis.

Relevant technical data such as operating hours, cycle counts or voltages are periodically monitored via a secure connection to our data center. The ZEISS Predictive Service application evaluates the performance of your microscope as system data can be received and analyzed.

Our support engineers will diagnose any issues by analyzing data on the Enterprise Server – remotely and without interruption to your operation.



- Maintain highest system availability
 Increase your uptime through close monitoring
 of the system's condition as remote support
 - of the system's condition as remote support can often provide immediate solutions
- Data security

Ensure highest data security standards using well established technologies like PTC Thingworx and Microsoft Azure Cloud. No personal or image data is uploaded, only machine data

■ Fast and competent support
Use secure remote desktop sharing to easily

get an expert connected

Optimum instrument performance
 As the status of your system is monitor.

As the status of your system is monitored, necessary actions can be planned before they become urgent

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| Typical Applications | Task | ZEISS Celldiscoverer 7 Offers |
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| Multiwell plates for Live Cell or fixed | Evaluate and document cell culture from multiwell plates. Scan the maximum area of a multiwell plate at different magnifications and resolutions. | Transmitted light – phase gradient contrast for high-resolution images through glass and plastic vessels |
| endpoint assays | | Up to 7 LED excitation wavelengths |
| | | Low magnification, large field of view – high numerical aperture lenses |
| | | Automatic sample carrier detection and calibration |
| | | Adaptive Lens Guard and automatic sample carrier calibration ensure maximized scan area depending on the plate type 100 % plate scanning from 2.5x to 100x is possible whole well – single shot |
| Label free assays | Perform label free growth curve assays over several days. | Transmitted light source: high-speed IR-LED (725 nm) offering low phototoxicity |
| | | Stable Incubation with temperature (heating/cooling), CO ₂ and O ₂ control |
| | | Simple and reproducible Hardware Autofocus for focus drift compensation |
| | | Autoimmerson for water immersion lens |
| High-Content Screening | cell culture from multiwell plates quickly. | Up to 7 LED excitation wavelengths |
| | | Autocorr objectives for automated aberration correction |
| | | Adaptive Lens Guard and automatic sample carrier calibration ensure maximized Scan area |
| | | Barcode reader for easy sample identificiation |
| | | Preview Scan |
| | | Open Application Developement for Python scripting – open access to third party analysis tools |
| | | Fast Multibandpass Main Beam Splitter and Emmission Filter Wheels |
| | | Large working distance enables higher/better 3D content screening |
| | Pharmacological or chemical or drug screening. | Option to add a plate loader |
| Fransfected and non-modified | Evaluate and document transfection rate and | Transmitted light – phase gradient contrast for high-resolution images through glass and plastic vessels |
| Live Cell Cultures | | Stable Temperature and O ₂ /CO ₂ controlled enviroment |
| | | Autoimmerson for water immersion lens |
| | Work with different sample carriers. | Automatic measurement of sample carrier bottom thickness and Autocorr Objectives for enhanced contrast and resolution |
| | | Adaptive Lens Guard and automatic sample carrier calibration ensure maximized scan area |

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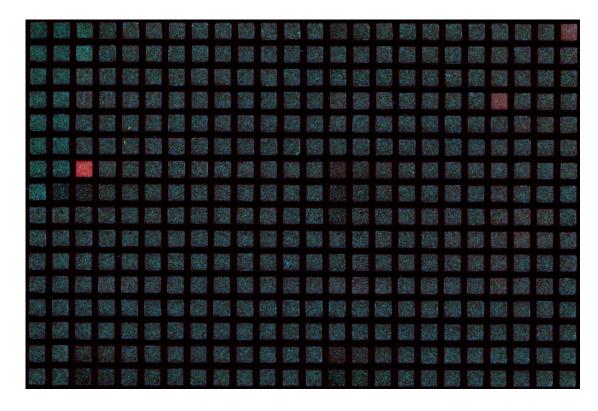
| Typical Applications | Task | ZEISS Celldiscoverer 7 Offers |
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| Label-free fixed and thin tissue slices or small organisms | Document and evaluate cell and tissue morphology and growth state. | Transmitted light – phase gradient contrast for high-resolution images through glass and plastic vessels |
| | Change quickly between large overview scans and high resolution imaging. | Quick change of field of view using triple magnification changer |
| | | Large working distances of 5× and 20×/0.7 objectives offer fast, high resolution and deep imaging |
| Fixed fluorescently labelled tissue, | Identification, quantification and qualification of cell types, pathological and pharmacological pathways using cell-, tissue and protein markers in 2D and 3D samples. | Up to 7 LED excitation wavelengths |
| cell culture samples or small organisms | | GPU-accelerated 3D-Deconvolution |
| | | Large working distances of 5× and 20×/0.7 objectives offer fast, high resolution and deep imaging |
| Multi-labelled living tissue section, organs, | . , , | Autoimmerson for water immersion lense |
| small organisms, organotypic-, spheriod or | | Autocorr objectives for automated aberration correction |
| cell culture preparations | | Stable incubation with temperature (heating/cooling), CO ₂ and O ₂ control |
| | | LED illumination unit with up to 7 excitation wavelengths |
| | | Experiment Feedback for adaptive experiments |
| | | GPU-accelerated 3D-Deconvolution |
| | | Large working distances of 5× and 20×/0.7 objectives offer fast, high resolution and deep imaging |
| | | Large working distances of 5× and 20×/0.7 objectives offer fast, high resolution and deep imaging |
| | | GPU-accelerated 3D-Deconvolution |
| Stimulus-induced responses of cells, | Observation of stimulus-induced responses of cells, tissue or organisms without disturbing the environmental control. | Semi-automatic dispensing work flow |
| tissue or whole organisms | | Dispensing unit allows to add compounds into the field of view |
| | | Option to install a perfusion chamber |

Tailored Precisely to Your Applications

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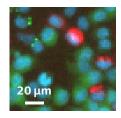
| Typical Applications, Typical Samples | Task | ZEISS Celldiscoverer 7 with LSM 900 Offers |
|--|---|--|
| Antibody stained tissue slices | Document morphological relations of structures | Airyscan 2 with GaAsP detector for imaging |
| Live cell culture | Study the motility of vesicles and organelles | Up to 8 frames per second confocal time lapse imaging |
| Live cell culture with two labels | Study the motility of subcellular structures | Airyscan 2 with GaAsP detector and Multiplex mode for time lapse imaging in 2D or 3D at up to 8 frames per second |
| | Explore the interaction of two proteins exploiting the Förster Resonance Energy Transfer effect | FRET analysis tool, available in ZEN (black edition) |
| Live cells with multiple labels | Image over a long time in an automated way | Experiment Designer software tool combined with three parallel spectral channels |
| Live or fixed cells with multiple labels and overlapping emission signals | Examine the interplay of multiple proteins | Parallel acquisition of all signals with three spectral channels and linear unmixing |
| Cellular structures with weak labels | Image subcellular structures at physiological expression levels | LSM 900 with GaAsP detector or Airyscan 2 |
| Study molecular dynamics | Photomanipulation | FRAP analysis tool, available in ZEN (black edition), classical timed bleaching or flexible interactive bleaching strategies |
| Plant roots | Follow the changes of subcellular structures over time with high resolution | Airyscan 2 with GaAsP detector for high resolution imaging beyond 40 μ m deep into tissue with up to 6 full frames per second (512 \times 512 pixel) |
| Model organisms, e.g. Zebrafish, <i>Drosophila</i> or <i>C. elegans, Arabidopsis</i> | See fine details of the organization and dynamics of endogeneously expressed FP proteins | Airyscan with GaAsP detector for high sensitivity imaging and increased resolution beyond 40 µm deep into tissue. |
| Cleared samples | Image whole organs or entire organisms | Specialized objective with long working distance and autocorrection for bottom material and thickness available (20×0.7) |

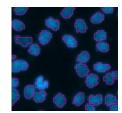
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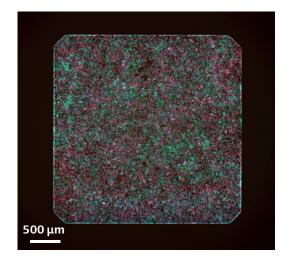


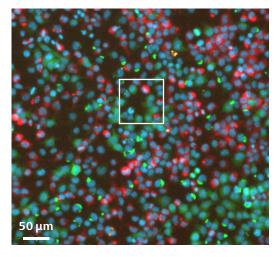
Whole well, single shot.

384 microwell plate imaged with 2.5x magnification in 3 channels. Each well fits into one single image. You avoid time-consuming scanning of wells and subsequent stitching and increase your throughput. He overall image quality and resolution allows e.g. segmentation of single cell nuclei and therefore counting of cells.



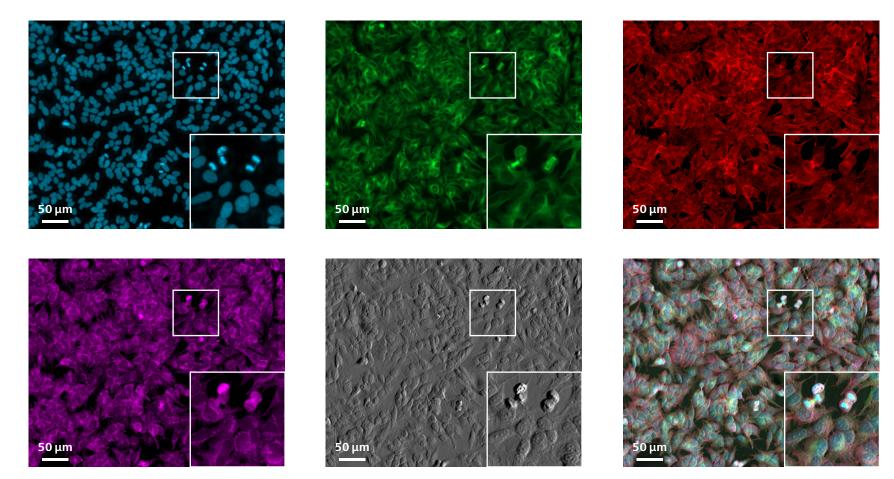






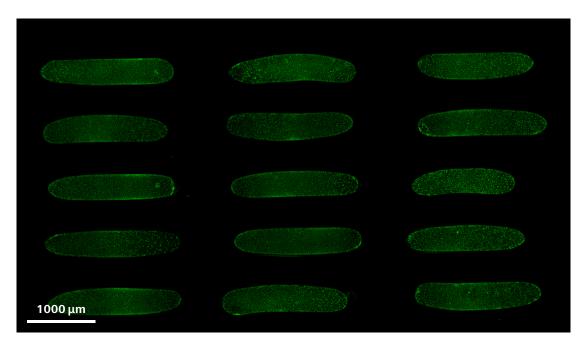
Sample courtesy of P. Denner, Core Research Facilities, German Center of Neurodegenerative Diseases (DZNE), Bonn, Germany.

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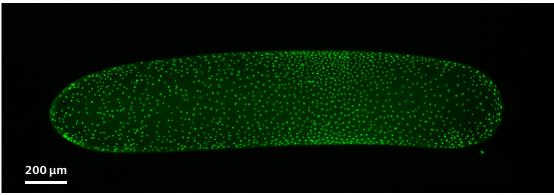


SH-SY5Y cells cultured on a 384 microwell plate. Five channel image at a single position using Plan-APOCHROMAT 20×/0.95; EDF from Z-stack; Hoechst-Chromatin (blue), anti-alpha-tubulin antibody FITC for alpha tubulin (green), Phalloidine for actin (red), MitoTracker deepRed for mitochondria (purple), phase gradient contrast, overlay image. Sample courtesy of P. Denner, Core Research Facilities, German Center of Neurodegenerative Diseases (DZNE), Bonn, Germany.

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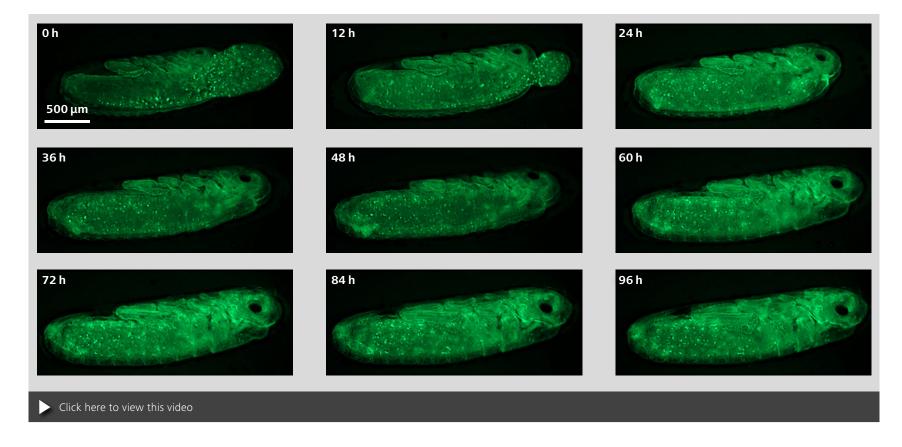


15 (out of 24) living cricket embryos mounted in low-melt agarose. Cells are expressing nuclear-localized GFP. The overview image shows a multiposition experiment. At each position two embryos fit into the field of view. Acquired within 30 seconds incl. Z-stacks of 17 images each (thickness 350 µm, 2.3 seconds). This enables imaging of multiple crickets in a synchronized way. The resulting spatio-temporal image resolution allows characterization of movement and division of single cells throughout the embryo during development. Magnification: 2.5× using short exposure times of 35 ms.



Sample courtesy of S. Donoughe, Biological Labs, Harvard University, Cambridge, USA

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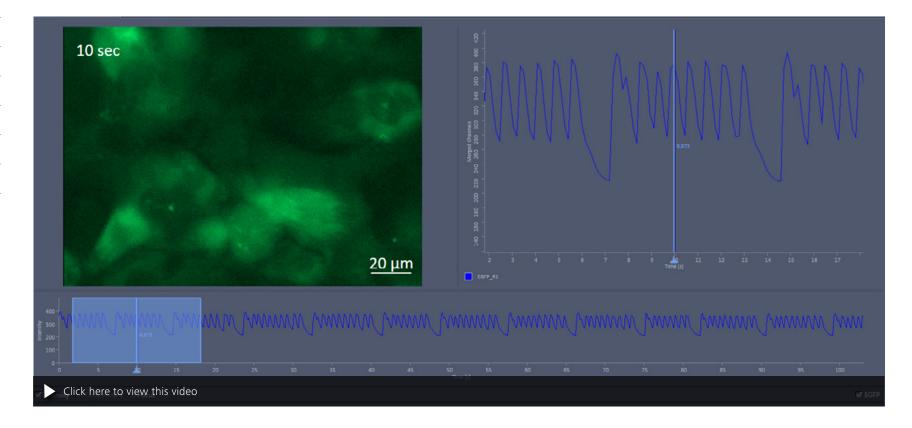


Five days long-term imaging of cricket embryogenesis. The development of an eGFP-expressing cricket embryo mounted in low-melt agarose was imaged every 5 minutes for a total length of 5 days. During the first day the retraction of the yolk and dorsal closure can be seen followed by further growing of the embryo. EDF-images created from Z-stacks; acquired with $2.5\times$ magnification using short exposure times of 35 ms.

Z-stacks were 350 μm thick and were acquired within 2.3 seconds.

Sample courtesy of S. Donoughe, BioLabs Building 2087, Harvard University, Cambridge, USA

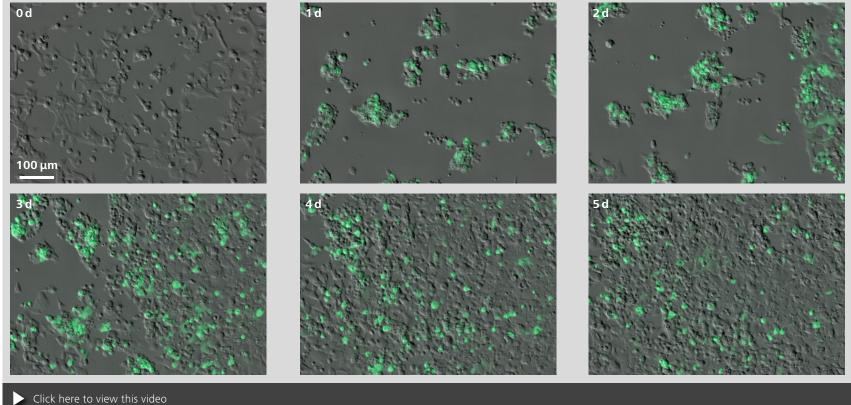
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Calcium imaging in beating cardiomyocytes stained in green using a Calcium kit; imaging with 8 fps using Plan-APOCHROMAT 50×/1.2 W with Autoimmersion; the green fluorescence changes intensity upon contraction of the cells; frequency of individual contractions analyzed with ZEN MeanROI tool; diagram shows delayed contraction in regular intervals caused by component given to the cells.

Sample courtesy of Sanofi-Aventis Deutschland GmbH, R&D IDD / in vitro Biology, Frankfurt, Germany

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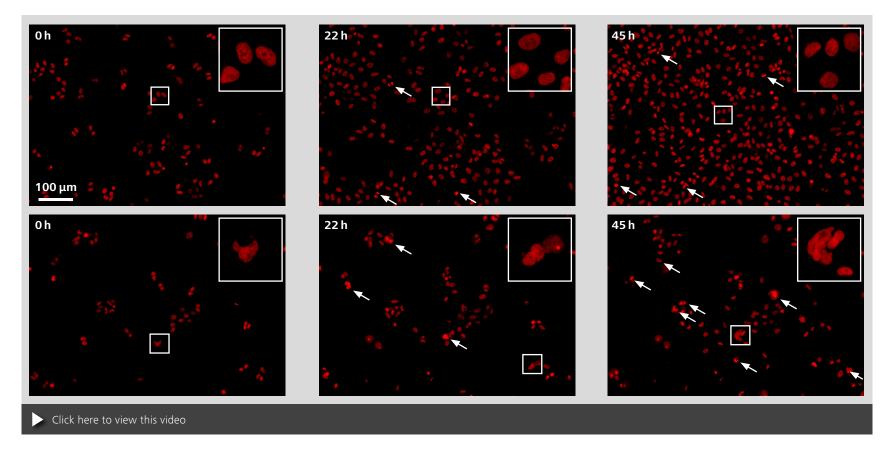


GFP HEK (Human Embryonic Kidney) cells, transiently expressing eGFP. Imaged through a 1 mm plastic bottom; images taken every 5 minutes for a total of 5 days; imaging started shortly after induction of the expression via Tetracyclin treatment. Overlay of phase gradient contrast and green (eGFP) fluorescence:

- After one day: cells are subconfluent and start to express eGFP. Due to the transient transfection and the Tetracyclin treatment some round and dead cells are visible.
- After two days: cells have recovered from the transfection and start to grow again.
- At the end of the time series: cells are confluent and bright green due to eGFP expression.

Sample courtesy of Sanofi-Aventis Deutschland GmbH; R&D IDD / in vitro Biology, Frankfurt, Germany

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48 h Cell Proliferation Assay Control vs. AuroroB Kinase siRNA Knockdown

HeLa Kyoto cells (Neumann et. Al., Nature 2010 Apr.1.; 464(7289):721-7) expressing H2B-mCherry were imaged every 30 minutes for 48 hours in a 96 well plate using Plan-APOCHROMAT 10×/0.5.

Top row: A series of images showing untreated control cells. The lack of dead cells and the healthy shape of the nuclei (arrows indicate methodic cells) clearly demonstrates the stability and homogeneity of the incubation, the stable focus, low phototoxicity as well as virtually no photobleaching.

Bottom Row: A series of images showing cells treated 24 h before acquisition with a siRNA against AuroraB Kinase on the same plate as the control (top row). The slower proliferation and the misshaped nuclei (arrows and insets) demonstrate the mitotic defects caused by the knockdown.

Sample courtesy of S. Reither, Advanced Light Microscopy Facility, EMBL, Heidelberg, Germany

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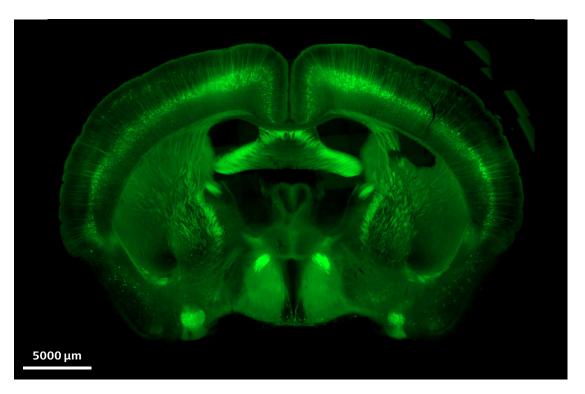
Expansion Microscopy in Mouse Brain

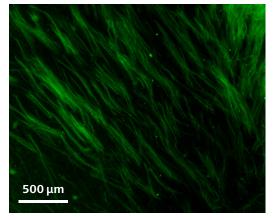
The goal of Expansion Microscopy is to make small structures visible that could otherwise not be observed with conventional or superresolution microscopy. Here, a protein-retention expansion technique was applied to expand the tissue. The sample is enlarged by a factor of 4.5 to 5- up to several mm in X/Y dimensions and several hundred μm in the Z dimension. Especially the $5\times/0.35$ and the $20\times/0.7$ objectives of Celldiscoverer 7 are well suited to image such samples as they have a large field of view, high resolution and a large working distance.

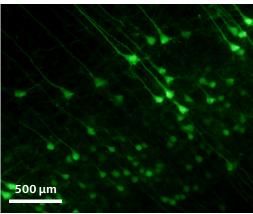
Top: Whole brain

Bottom left: Axon bundles Bottom right: Pyramidal cells

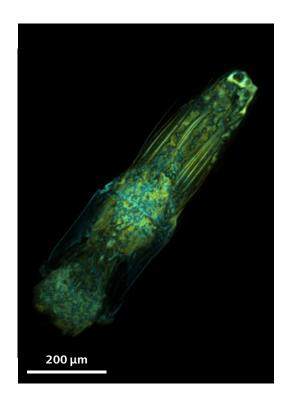
The images shown here are extended-depth-offocus images created from Z-stacks acquired with a 2.5x magnification imaged through 1.2 mm of polystyrene. Staining: YFP expressing neurons.







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Autofluorescence Imaging of Arachnids

Small Arachnids where collected from tropical leaves in South America. Imaging with Celldiscoverer 7 saves time, since the low magnification objectives $(5\times/0.35 \text{ and } 20\times/0.7)$ deliver finest details in large fields of view.

A combination of several wavelengths was used to observe autofluorescence. The images shown here are extended-depth-of-focus images created from Z-stacks.

Left: Genital of the third leg of Huitaca sp. imaged with a $20\times$ magnification.

Center: Same as before but excited with a different combination of wavelengths.

Right: *Microgavia oviformis* imaged with 2.5× magnification.

Sample courtesy of L. Benavides, Giribet Lab, Harvard University, Cambridge, USA

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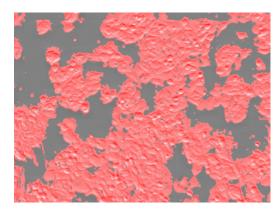
Application for Label-Free Measurement of Cell Proliferation

The growth of cultured cells has been imaged in long-term time-lapse movies over 72 hours using phase gradient contrast (image 1).

To quantify proliferation, cell region (image 2, red overlay) was detected automatically using supervised machine learning (random forests) in each time frame.

The growth curve (image 3) shows the relative cell coverage over time, averaged for all images in one well. The assay allows image based cell proliferation measurements.

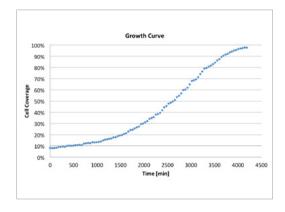
By using label-free imaging in phase gradient contrast, cell growth is not affected by phototoxicity or any further sample processing.



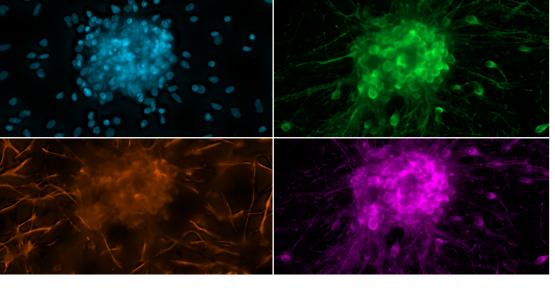
Sample and assay courtesy of P. Denner, Core Research Facilities, German Center of Neurodegenerative Diseases (DZNE), Bonn, Germany.

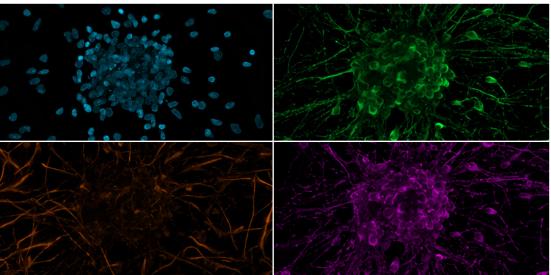
This approach offers several advantages:

- Very low disturbance, non-invasive monitoring of cells
- Kinetic live cell data, no single end point.
- Compatible to standard micro-well plates (e.g. 96well or 384well).
- Applicable for screening cell-based applications.



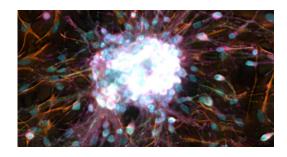
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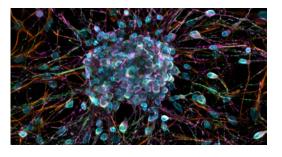
Rat cortical primary neuron culture. Antibody staining of bIII-tubulin (Cy2, green), Nestin (Cy3, red) and DCX (Cy5, purple), nuclei stained with DAPI (blue). Maximum intensity projection of a Z-stack.

Top row: Conventional widefield images.

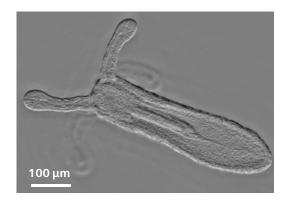


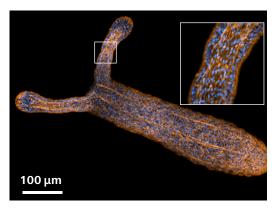
Bottom row: Deconvolved images using GPU-based deconvolution. Deconvolution algorithm: constrained iterative using a depth variant point-spread function.

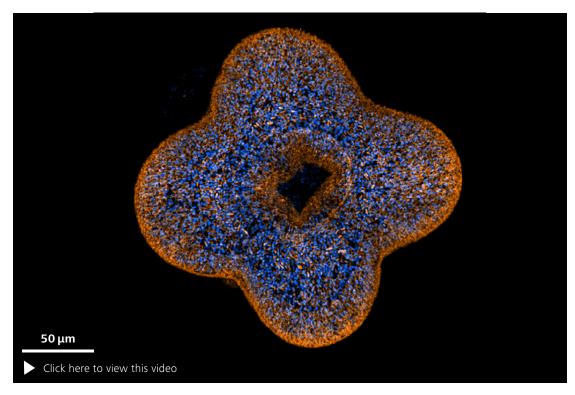
Sample courtesy of H. Braun, LSM Bioanalytik GmbH, Magdeburg, Germany.



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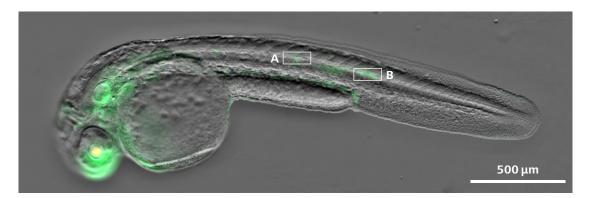
Sample courtesy of A. Stokkermans, Ikmi Group, EMBL, Heidelberg, Germany

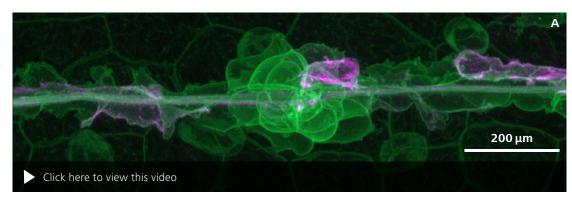
Fixed starlet sea anemone (Nematostella vectensis) stained with Hoechst (nuclei) and Phalloidin (actin). Side view imaged with a combination of camera based phase gradient contrast mode (top) and high sensitivity mode with Airyscan 2 (bottom). Maximum intensity projection of 19 z-planes.

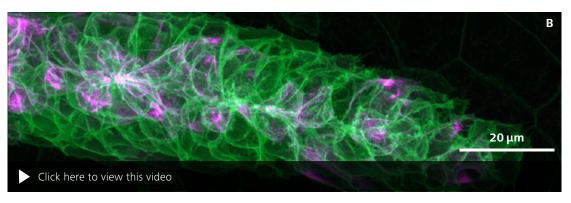
Fine image details and high signal to noise ratio can clearly be seen on the insert in the top right image, showing an enlarged view of a tentacle area.

Video: Top view of a young animal, showing mouth and four tentacle buds. Maximum intensity projection of 69 z planes imaged with Airyscan 2 Multiplex. Images were acquired using the water immersion objective with a total magnication of 25× and a numerical aperture of 1.2.

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Lateral line primordium migration and deposition of immature neuromasts in a Zebrafish embryo (*Danio rerio*). Animals were anesthetized and embedded using low concentrated agarose in a glass bottom petridish.

Initial camera based imaging allowed for a quick and easy sample navigation (top) combining Phase Gradient Contrast with fluorescence acquisition.

Subsequent high resolution imaging with Airyscan 2 in Multiplex mode was done on individual positions identified in the widefield image (white boxes).

- A) Maximum intensity projections of an immature neuromast (127 z planes).
- B) Maximum intensity projections of the lateral line primordium tip migrating through the animal (155 z-planes).

Green: LYN-eGFP (mebranes);

Red: tagRFP-T-UTRCH (actin).

The gentle and fast image acquisition that is inherent to the Airyscan 2 Multiplex mode is very benificial for this kind of application. The animal is unperturbed by the imaging while images with a very high signal to noise ratio as well as level of detail can be acquired at the same time.

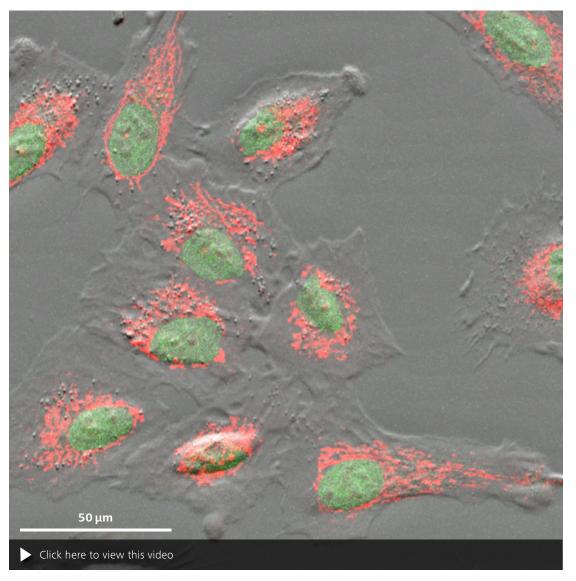
Sample courtesy of J. Hartmann and D. Gilmour, EMBL, Heidelberg, Germany

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Primary lung fibroblasts stained with mitotracker red (mitochondria) and a DNA marker (nuclei).

The acquisition seemlessly combines two imaging modes – the fluorescent channels where captured in confocal mode using highly sensitive GaAsP detectors while the Phase Gradient Contrast is camera based.

A timelapse of 2.5 h was acquired using a $40 \times$ magnification with a numerical aperture of 0.95.



Sample courtesy of A. Hocke, Charité, Berlin, Germany

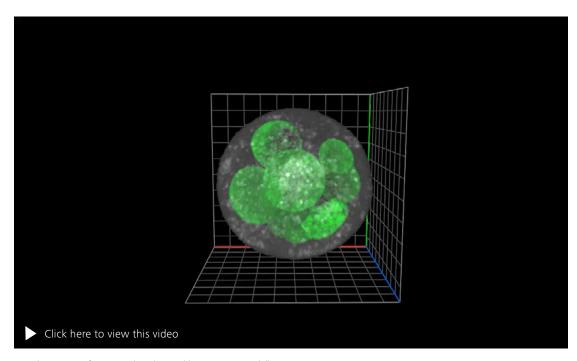
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Organoid from a human breast cancer cell line. The cells express GFP-labeled H2B (nucelei) and mCherry (cytoplasmic staining depicted here in grey for better visualization).

Several organoids where grown in a multiwell plate with Matrigel. Initial sample navigation was performed using the transmitted light at a low magnification of 2.5× to identify interesting organoids.

Subsequently, high resolution images were acquired using the water immersion objective with a total magnification of 50×. 61 z-planes were acquired using ZEISS Celldiscoverer 7 with LSM 900 and Airyscan 2 in Multiplex mode.

One can clearly appreciate the robustness of the imaging given that Matrigel is not an ideal optical medium and the organoid was imaged at a distance of several micrometers from the coverglass.

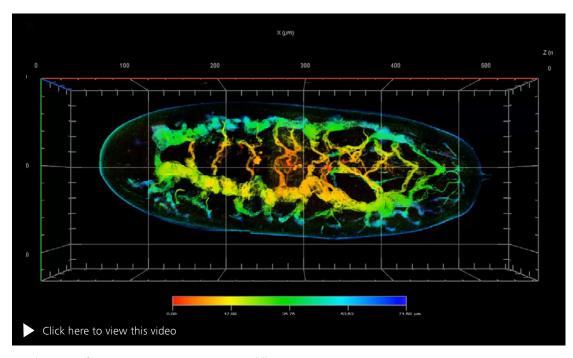


Sample courtesy of S. Gawrzak and M. Jechlinger, EMBL, Heidelberg, Germany

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Trachea system in a living fruitfly embryo (*Drosophila melanogaster*) imaged using ZEISS Celldiscoverer 7 with LSM 900 and Airyscan 2 in Multiplex mode. A water immersion objective with a magnification of 25× and numerical aperture of 1.2. in combination with multi-tile acquisition (8 tiles, 143 z-planes) was used.

CD4-mIFP under a tracheal promoter color coded for depth.



Sample courtesy of D. Rios-Barrera, Leptin Group, EMBL, Heidelberg, Germany

ZEISS Celldiscoverer 7 at Work

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Caenorhabditis elegans germline. Decapitated nematodes where localized in widefield mode using a low magnification of 2.5× (transmitted light and fluorescence, DAPI; left). This allowed for an easy and convenient workflow to identify areas of interest for subsequent fast high resolution imaging in Multiplex mode for ZEISS Celldiscoverer 7 with LSM 900 and Airyscan 2 (right). A 25× magnifaction with water immersion and NA 1.2 was used to generate a z-stack of 62 planes.

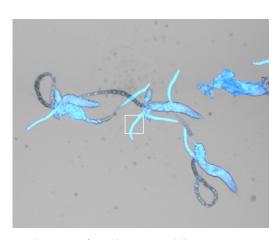
Individual chromosomes in different meiotic cells are clearly distinguishable – see magnified box.

Blue: DAPI (DNA);

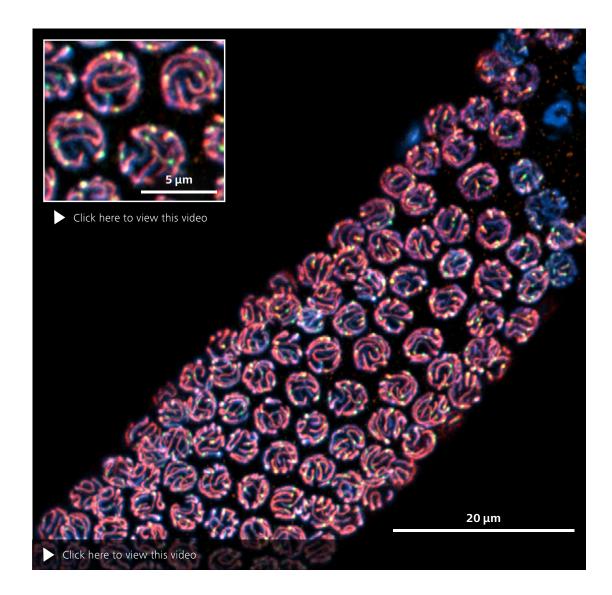
Green: Alexa 488 (cross-over sites);

Orange: Alexa 546 (synaptonemal complex);

Red: Alexa 647 (chromosome axis).



Sample courtesy of S. Köhler, EMBL, Heidelberg, Germany



Your Flexible Choice of Components

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1 Microscope

- ZEISS Celldiscoverer 7
- Automatic sample container recognition
- Barcode reader
- Focus stabilization
- Magnification changer $0.5 \times /1 \times /2 \times$
- Apochromatic FL beampath with adaptive field stop
- ZEISS Axiocam 506 mono or Axiocam 512 mono
- Additional camera port
- On-axis access for dispensing
- UV-disinfection

2 Objectives

- Plan-APOCHROMAT 5×/0.35
- Plan-APOCHROMAT 20×/0.7 autocorr
- Plan-APOCHROMAT 20×/0.95 autocorr
- Plan-APOCHROMAT 50x/1.2 W autocorr autoimmersion

3 Illumination

- Transmitted light unit:
 IR-LED (725 nm) brightfield, oblique contrast,
 phase gradient contrast
- Fluorescence:
 LEDs 385, 420, 470, 520, 567, 590 and 625 nm
 High-efficiency multibandpass filter sets
 Additional emission filter wheel

4 Imaging Systems

■ LSM 900 with Airyscan 2

5 Accessories

- Temperature and atmospheric control (heating/cooling; CO₂, O₂)
- Insert plates and perfusion chambers for dishes, multi-chamber slides and standard slides

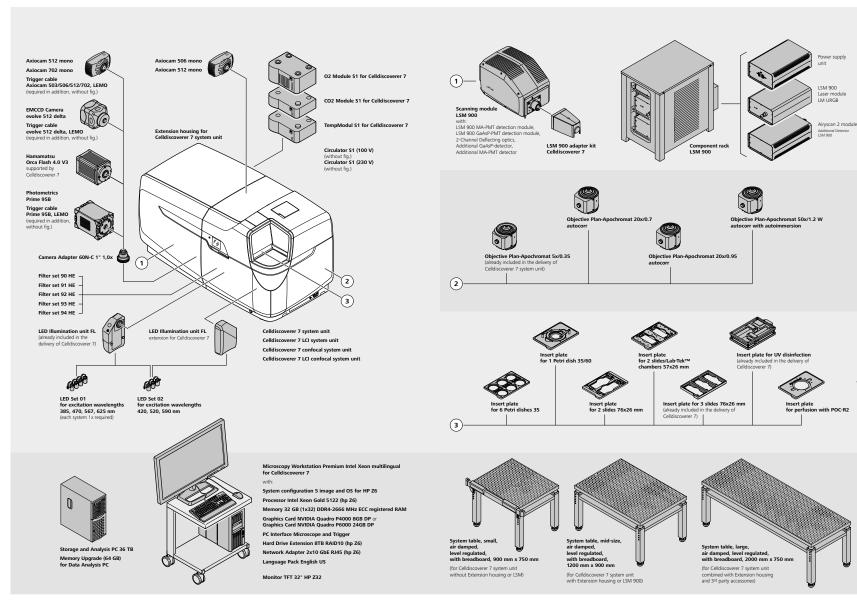
- Additional recommended cameras
 - ZEISS Axiocam 512 mono
 - ZEISS Axiocam 702 mono
 - Photometrics EMCCD evolve 512 delta
 - Hamamatsu Orca Flash 4.0
 - Photometrics Prime 95B

6 Software

- ZEN celldiscoverer includes modules for multidimensional image acqusition, Tiles & Positions, Experiment Designer, advanced image processing and analysis tools
- Recommended additional modules:
 - GPU-based deconvolution (GPU-DCV)
 - 3Dxl Viewer powered by arivis®
 - Open application development (OAD)

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| Dimensions | Width (approx.) | Depth (approx.) | Height (approx.) | Weight (approx.) |
|-----------------------------------|-----------------|-----------------|------------------|------------------|
| Celldiscoverer 7 | 710 mm | 640 mm | 700 mm | 136 kg |
| Footprint Celldiscoverer 7 | 585 mm | 560 mm | | |
| Incl. Extension housing | 1270 mm | 640 mm | 700 mm | 187 kg |
| Footprint incl. Extension housing | 1170 mm | 560 mm | | |
| Celldiscoverer 7 incl. LSM 900 | 1310 mm | 690 mm | 705 mm | |
| Component rack | 400 mm | 550 mm | 600 mm | 35 kg |
| Airyscan 2 | 400 mm | 250 mm | 145 mm | 5 kg |
| Power Supply | 400 mm | 250 mm | 145 mm | 6 kg |
| Laser module | 400 mm | 250 mm | 145 mm | 10 kg |

| Technical data |
|------------------|
| Celldiscoverer 7 |

| Celldiscoverer 7 and Extension housing | Noise emission | According to EN 55011 class A | | |
|--|---|---|--|--|
| | Noise immunity | According to DIN EN 61326-1 | | |
| | Protection class | 1 IP 20 To EN 55011 Class A Closed room facility To DIN EN 61010-1 (IEC 61010-1) conforming to CSA and UL regulations 2 | | |
| | Ingress protection rating | | | |
| | Radio interference suppression | | | |
| | Type of operating site | | | |
| | Electrical safety | | | |
| | Degree of pollution | | | |
| | Overvoltage category | II | | |
| Celldiscoverer 7 | Line input voltage; max. current | 100 V to 240 V ± 10 %; 6A~ | | |
| | Line frequency | 50 Hz – 60 Hz | | |
| Celldiscoverer 7 incl. LSM 900 / | Input for connection of Celldiscoverer 7 | 100 V to 240 V \pm 10 %, 50 Hz $-$ 60 Hz, max. 4.0 A \sim | | |
| Extension housing | Output to internal 6 sockets | 100 V to 240 V ± 10 %, 50 Hz – 60 Hz | | |
| | Permissible total current on 6 internal sockets | Max. 4.0 A~ | | |
| | | The internal sockets can be connected via the software | | |
| | | The extension housing is powered by Celldiscoverer 7 | | |

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| Storage (in packaging) | Permissible ambient temperature | +5°C to +40°C |
|---|--|---|
| | Permissible relative air humidity (no condensation) | max. 75 % at +35 °C |
| Transport (in packaging) | Permissible ambient temperature | −20 °C to +55 °C |
| | Permissible relative air humidity (no condensation) | max. 75 % at +35 °C |
| Operation | Permissible ambient temperature | +15°C to +35°C |
| | Recommended ambient temperature (e.g. for incubation) | +18°C to +25°C, optimally +22°C |
| | Warm-up time | 1 h for standard imaging; ≥4 h for high-precision and/or long-term measurements |
| | Permissible relative air humidity | max. 65 % at 30 °C |
| | Atmospheric pressure | 800 hPa to 1060 hPa |
| XYZ motorization Motorized xy-scanning stage | Travelling range | 300 mm × 140 mm |
| Motorized xy-scanning stage | Travelling range | 300 mm × 140 mm |
| | Reproducibility | ± 1 μm |
| | Absolute precision | ± 5 µm |
| | Resolution | 0.1 μm |
| Motorized z-drive | Reproducibility | ± 0.025 μm |
| | Absolute precision | 0.14 µm |
| | Resolution | ± 0.01 µm |
| | | |
| Optical specifications | | |
| Nosepiece | 4x motorized nosepiece in combination with the 3x magnificati functionality of 12 objectives | ion changer this offers the |
| Magnification changer, afocal | 0.5x, 1x, 2x magnification, providing t for each objective depending on the objective configurati range from 2.5x - 100x switching between magnifications ~1 s enables constant working distances for | ion it offers a magnification |

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| | | 1 | Magnification cha | inger | Auto- | Auto- | Temperature | Thick vessel bottom up to | Thin vessel bottom 0.13 – 0.21 mm glass/COC ¹ | Working |
|---|--|---------------------|-------------------|-------------------------------|------------|-----------|----------------|---------------------------|--|---------|
| | | 0.5× | 1× | 2× | correction | immersion | control | 1.2 mm PS ² | 0.15-0.21 mm PS ² | distanc |
| Plan-Apochromat 5×/0.35 | • | M = 2.5 NA = 0.1 | | $M = 10 \times $ NA = 0.35 | _ | - | • | • | • | 5.10 mr |
| Plan-Apochromat 20×/0.7 autocorr | 0 | M = 10x NA = 0.3 | | $M = 40 \times $ $NA = 0.7$ | • | - | • | • | • | 2.20 mr |
| Plan-Apochromat 20×/0.95 autocorr | 0 | M = 10x NA = 0.5 | | $M = 40 \times $ NA = 0.95 | • | - | • | - | • | 0.76 mr |
| Plan-Apochromat 50×/1.2 W autocorr, autoimm. | 0 | M = 25 NA = 1.2 | | M = 100× NA = 1.2 | • | • | • | - | • | 0.84 mr |
| Adaptive Lens Guard | • | | cally maximizes s | - | - | | | vith other hardwa | ire or sample vessels | |
| Temperature control | all objectives are equipped with heating elements for temperature control in combination with the optional heating unit, objective temperature is adjusted automatically, depending on the user-defined sample temperature enables stable and homogeneous temperature within the sample chamber | | | | | | | | | |
| Adaptive Autocorr | automatic correction of aberrations (for high magnification objectives) adapts objectives automatically to vessel bottom material and thickness enables correction of aberration due to high penetration depths and refrective index mismatch of the sample (5× objective is not sensitive to variations of bottom thickness and material and does not require a correction) | | | | | | | | | |
| Autoimmersion, water | comes along with the Plan-Apochromat 50×/1.2 W objective enables automatic supply and removal of water immersion water level is automatically indicated in the control software and on the display upgradable in the field | | | | | | | | | |
| | resolution im | provement | | | | | | | | |
| Optional LSM 900 with Airyscan 2 for up to 1.5× i | | ochromat 5×/0.3 | 5 Plar | -Apochromat 2 | 20×/0.7 | Plan-Apo | ochromat 20×/0 | | n-Apochromat 50×/1. | 2 W |
| Objective | | | | | | | | 2 0 5 | 1 | |
| Objective Magnification changer | Plan-Ap | | 2× 0.5> | 1× | 2× | 0.5× | 1× | 2× 0.5 | × 1× | 2× |
| Optional LSM 900 with Airyscan 2 for up to 1.5× i Objective Magnification changer Usage with Airyscan MPLX | | | | 1× + | 2× | 0.5× + | 1× ++ | ++ ++ | × 1× | 2× |

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| Focus | |
|--|--|
| Hardware-based focus finder | automatically focusses on the sample (lower side of sample) a user-defined offset can be used to change the default position enables automatic generation of focus maps for microwell plates compatible with every objective and filter set can be combined with focus stabilization and ZEN blue software autofocus |
| Hardware-based focus stabillization | focus stabilization system maintains focus position over long-term compatible with every objective and filter set hardware and software support for multi-position and multi-offset stabilization can be combined with focus finder and ZEN blue software autofocus |
| Software-based autofocus | focusses automatically on user-defined structures and regions of interest based on the image content can be combined with focus finder and focus stabilization |
| Transmitted light and contrasting techniques | |
| Transmitted light unit | fully compatible with fluorescent applications, environmental control, dispensing and perfusion option enables label-free imaging or provides additional information in combination with fluorescent applications |
| Lightsource | ● ■ high-speed IR-LED (725 nm) offering low phototoxicity |
| Contrast techniques | brightfield oblique contrast adaptive phase gradient: adapts automatically to vessel geometry providing excellent contrast to the edges of the vessels all contrast techniques are compatible with all objectives, filter sets and sample vessels, i.e. plastic and glass incl. lids |

• Component always included O Component optionally available

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| Fluorescence illumination unit | apochromatic excitation beampath incl. adaptive field stop up to seven LEDs (385 / 420 / 470 / 520 / 567 / 590 / 625 nm) life time of LEDs >10,000 h switching between LEDs <1 ms |
|--|--|
| LEDs are synchronized with image acquisition | Sample is only exposed during image acquisition (acquisition trigger mode) thus reducing phototoxicity. |
| LEDs are synchronized with the live-window | Sample is only exposed during live-window update (live-window trigger mode), significantly reducing phototoxicity during sample navigation |
| Automated excitation field stop | A motorized field stop adapts automatically to the current field of view thus reducing phototoxicity effectively. |
| Switching time between FL channels | switching between fluorescence channels using high-efficient multi-bandpass filter sets <1 ms switching 5-position beamsplitter wheel <80 ms |
| 5-position beamsplitter wheel | 5× position beamsplitter wheel switching time <80ms |
| Emission filter wheel | 7× motorized emission filter wheel user accessible fits 25 mm emission filters switching emission filter wheel <80 ms |
| Filter sets | Filter set 90 HE quad-band filter set for 385 nm, 470 nm, 567 nm, 625 nm LED and IR-TL LED beamsplitter RQFT 405+493+575+653; emission filter QBP 425/30+514/30+592/25+709/100 additional band for transmitted light |
| | Filter set 91 HE triple-band filter set for 420 nm, 520 nm, 590 nm LED and IR-TL LED beamsplitter RTFT 450+538+610; emission filter TBP 467/24+555/25+687/145 additional band for transmitted light |
| | Filter set 92 HE triple filter set for 385 nm, 470 nm, 590 nm LED and IR-TL LED beamsplitter RTFT 405+493+610; emission filter TBP 425/30+524/50+688/145 additional band for transmitted light |
| | Filter set 93 HE double bandpass for 470 nm, 567 nm and IR-TL LED beamsplitter RDFT 493+575; emission filter TBP 514/32+605/50+730/60 additional band for transmitted light |
| | Filter set 94 HE double filter set for 385 nm, 520 nm and IR-TL LED beamsplitter RDFT 405+538; emission filter TBP 444/69+581/77+730/60 additional band for transmitted light |

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| BP 385/30 | | | Filter sets | 90 HE | 91 HE | 92 HE | 93 HE | 94 HE | |
|--|------------------------------|---------|-------------|-------|--------|-------------------|------------------|----------------|--|
| BP 385/30 | | | LEDs [nm] | quad | triple | triple | dual | dual | |
| LED 567 RP 469/38 X | LED Set 1 | | | × | | × | | × | DAPI, Hoechst 33342 & 33258, Alexa 350 & 405, ATTO 390, True Blue, EBFP, T-Sapphire CellTracker Blue, LysoTracker Blue, wtGFP (uv), Aminocoumarin, Cascade Yellow |
| LED 5et 2 LED 420 RP 423/44 X X X X X X X X X | | | | × | | × | × | | TagGFP, LysoTracker Green, ATTO 465, ATTO 490, Oregon Green Bapta, BOBO-1, Cytox |
| LED 525 BP 631/33 X Bodipy 630/650-X, Bodipy 650/665-X, CF™620R, CF™633, CF™640R, DyLight 649, P5mOrange (red.), IRFP70 LED 5et 2 LED 420 BP 423/44 X X Alexa Fluor 430, ECFP, ATTO 425, ATTO 430LS, SpectrumAqua, Cerulean, mCFP, CyPet, Y66W, mKeima-Red, LysoSensor™ Green DND-153, SYTOX Blue mycin A3, POPO-1, PO-PRO-1, SYTO 40, SYTO 41, SYTO 42, SYTO 43 LED 520 BP 511/44 X X X Alexa 514 8 532, eYFP, Calcein, Fluo-4, Fluo-8, Bodipy 515, YOPro-1, YOYo-1, Green, Syto 23, Thiazole Orange, LysoTracker® Green DND-26, mEos3.2 (gree mEOS2.0, mCitrine, mVenus, Topaz Alexa Fluor 594, Cy3.5, mPlum, mRaspberry, mNeptune, mCherry, pa-mRFP1, mEos2 (red.), mEos3.2 (red.), LipidTOX™ Red, Calcein red-orange, CellTracker Red, Tell-Tracker Red, CellTracker Red, Tell-Tracker Red, CellTracker Red, | | • | | × | | | × | | Cy3, Bodipy TMR, mBanana, mOrange, TurboRFP, tdTomato, TagRFP, DsRed2 ("RFP"), TRITC, PAmCherry, PATagRFP, Alexa Fluor 555 & 546, DsRed monomer, SNARF, PO-PRO-3, Magnesium Orange, SYTO 82 |
| P 423/44 | | | | × | | | | | Cy5, Alexa Fluor 610, 633, 635 & 647, ATTO 610 to 647N, ATTO Oxa12, ATTO Rho14, Bodipy 630/650-X, Bodipy 650/665-X, CF™620R, CF™633, CF™640R, DyLight 633, DyLight 649, PSmOrange (red), iRFP670 |
| Alexa Fluor 594, Cy3.5, mPlum, mRaspberry, mNeptune, mCherry, pa-mRFP1, mEos2 (red), mEos3.2 (gree mEOS2.0, mCitrine, mVenus, Topaz Alexa Fluor 594, Cy3.5, mPlum, mRaspberry, mNeptune, mCherry, pa-mRFP1, mEos2 (red), mEos3.2 (red), LipidTOX TM Red, Calcein red-orange, CellTracker Red, ER-Tracker Red, CellTrace BODIPY® TR TL IR Channel IR LED 725/50 X X X X X X All filter sets offer an IR transmitted light bandpass. This bandpass enables IR-brightfield contrast without switching any filter components and without affecting FL-efficiency. Lasers Laser module URGB (pigtailed; 405, 488, 561, 640 nm) O Single-mode polarization preserving fiber Typical total dynamic range of 10.000:1; direct modulation 500:1 Diode laser (405 nm, 5 mW) Diode laser (488 nm, 10 mW) | LED Set 2 | | | | × | | | | mCFP, CyPet, Y66W, mKeima-Red, LysoSensor™ Green DND-153, SYTOX Blue, Chromo |
| LED 590 BP 591/27 X X X MEos2 (red), nEos3.2 (red), LipidTOX ^M Red, Calcein red-orange, CellTracker Red, ER-Tracker Red, CellTrace BODIPY® TR TL IR Channel IR LED 725/50 X X X X X X X X IR-brightfield contrast without switching any filter components and without affecting FL-efficiency. Lasers Laser module URGB (pigtailed; 405, 488, 561, 640 nm) O Single-mode polarization preserving fiber Typical total dynamic range of 10.000:1; direct modulation 500:1 Diode laser (488 nm, 10 mW) Diode laser (488 nm, 10 mW) | | 0 | | | × | | | × | Alexa 514 & 532, eYFP, Calcein, Fluo-4, Fluo-8, Bodipy 515, YoPro-1, YoYo-1, Calcium Green, Syto 23, Thiazole Orange, LysoTracker® Green DND-26, mEos3.2 (green), mEOS2.0, mCitrine, mVenus, Topaz |
| Channel IR LED 725/50 X X X X X X X IR-brightfield contrast without switching any filter components and without affecting FL-efficiency. Lasers Laser module URGB (pigtailed; 405, 488, 561, 640 nm) O Single-mode polarization preserving fiber Typical total dynamic range of 10.000:1; direct modulation 500:1 Diode laser (405 nm, 5 mW) Diode laser (488 nm, 10 mW) | | | | | × | × | | | · · · · · · · · · · · · · · · · · · · |
| Laser module URGB (pigtailed; 405, 488, 561, 640 nm) Single-mode polarization preserving fiber Typical total dynamic range of 10.000:1; direct modulation 500:1 Diode laser (405 nm, 5 mW) Diode laser (488 nm, 10 mW) | | • | | × | × | × | × | × | IR-brightfield contrast without switching any filter components and without |
| Laser module URGB (pigtailed; 405, 488, 561, 640 nm) Single-mode polarization preserving fiber Typical total dynamic range of 10.000:1; direct modulation 500:1 Diode laser (405 nm, 5 mW) Diode laser (488 nm, 10 mW) | | | - | | | | | | |
| (pigtailed; 405, 488, 561, 640 nm) Typical total dynamic range of 10.000:1; direct modulation 500:1 Diode laser (405 nm, 5 mW) Diode laser (488 nm, 10 mW) | Lasers | | | | | | | | |
| Diode laser (488 nm, 10 mW) | Laser module URGB | | | | Sin | gle-mode polar | ization preservi | ng fiber | |
| Diode laser (488 nm, 10 mW) | (pigtailed; 405, 488, 561, 6 | 540 nm) | | | Тур | oical total dynan | nic range of 10 | .000:1; direct | t modulation 500:1 |
| Diode laser (488 nm, 10 mW) | | | | | | ode laser (405 n | m, 5 mW) | | |
| Diode (SHG) laser (561 nm, 10 mW) | | | | | | ode laser (488 n | m, 10 mW) | | |
| | | | | | Dic | ode (SHG) laser | (561 nm, 10 m) | W) | |
| Diode laser (640 nm, 5 mW) | | | | | Dic | ode laser (640 n | m. 5 mW) | | |

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| Sample mounting | | |
|--|---|--|
| Insert plate for 1 Petri dish 35/60 | 0 | for mounting of Petri dishes fits one Petri dish d = 35 mm or d = 60 mm, autoclavable |
| Insert plate for 6 Petri dishes 35 | 0 | for mounting of Petri dishes fits six Petri dishes d = 35 mm, autoclavable |
| Insert plate for 2 slides 76×26 mm | 0 | ■ for mounting of slides ■ fits two slides 76 × 26 mm, autoclavable |
| Insert plate for 3 slides 76×26 mm | 0 | ■ for mounting of slides ■ fits three slides 76 × 26 mm, autoclavable |
| Insert plate for 2 slides/Lab-Tek™ chambers 57×26 mm | 0 | ■ fits two Lab-Tek™ chambers 57 × 26 mm, autoclavable |
| Insert plate for perfusion with POC-R2 | 0 | ■ fits for perfusion with POC-R2 |
| Internal camera* | • | Axiocam 506 mono, Axiocam 512 mono |
| Detection options | | |
| External camera port ** | • | external, user accessible camera port to mount additional cameras |
| | | ■ motorized switching between internal and external camera <200 ms |
| Additional/optional cameras | 0 | Axiocam 512 mono |
| | 0 | Axiocam 702 mono |
| | 0 | Photometrics EMCCD evolve 512 delta |
| | 0 | Hamamatsu Orca Flash 4.0 |
| | | |
| | 0 | Photometrics Prime 95B |
| LSM 900 | 0 | Photometrics Prime 95B two spectral detection channels, GaAsP (typical QE 45 %) or multialkali (MA) PMT (typical QE 25 %) |
| LSM 900 | | |

• Component always included O Component optionally available * Select one internal camera ** Not available on systems with LSM 900

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| Resolution and speed (examples) | |
|--------------------------------------|--|
| Pixel resolution | depending on the magnification and camera: 1.82 µm @ 2.5x using Axiocam 506 0.23 µm @ 20x with Axiocam 506 1.24 µm @ 2.5x using Axiocam 512 0.03 µm @ 100x using Axiocam 512 |
| Typical scan speeds | 96 well plate, four channels, exposure 50 ms per channel, full resolution, one position per well: <4 min 96 well plate, three confocal channels simultaneously (multicolor track), image size 512 x 512 px, bidirectional scan at max. speed, one position per well: <2.5 min. (with optional LSM 900) 384 well plate, single channel, exposure 100 ms, full resolution, 1 position per well (e.g. whole well single shot): < 6 min 384 well plate, whole well using a high resolution 20x objective, four channels, exposure 50 ms per channel, full resolution: <2.5 min |
| Automatic sample recognition | |
| Pre-scan unit (incl. barcode reader) | automatically detects vessel types before final mounting: slides Petri dishes (35/60 mm) LabTek-chamber slides (incl. number of wells) microwell plates incl. plate type, i.e. number of wells The following 1D barcodes are detected on slides and wellplates: Code 39 (3of9 und W/MOD43) Code128 Auto, Code128 A, Code128 B, Code128 C Interleaved 2of5 UPC A und UPC E EAN 8 und EAN 13 Codebar UCC/EAN 128 on slides the following 2D barcodes are detected: DataMatrix QR-Code |
| Automatic vessel bottom recognition | automatic detection of vessel bottom material (glass/COC¹ and PS²) automatically adjusts autocorr objectives to the material automatic detection of vessel bottom thickness automatically adjusts autocorr objectives to the thickness automatically measures vessel skirt height, e.g. the distance between the support area and the actual sample bottom delivers the skirt height to the Adaptive Lens Guard to update the scanning area |
| Automatic plate calibration | automatically calibrates individual plates, i.e. well diameter and distance, plate length, height and rotation |

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| Environmental control | |
|----------------------------------|---|
| TempModule S1 | controls temperature of bottom and top plate of sample chamber temperature range within sample chamber: 30 – 45 °C temperature homogenity across a whole microwell plate: ± 0.6 @ 37 °C operated by ZEN blue control software |
| CO ₂ Module S1 | generates a stable, user defined CO₂ concentration within the sample chamber ensures an optimal and stable pH value in cell culture media over long term a built-in CO₂ sensor permanently monitors the CO₂ concentration operated by ZEN blue control software |
| O ₂ Module S1 | O₂-control device to achieve a stable, controlled decrease of the O₂ concentration by displacement with N₂ within the sample chamber a built-in O₂ sensor permanently monitors the O₂ concentration. operated by ZEN blue control software |
| Humidifier unit | o prevents evaporation of culture medium during long-term experiments liquid level is indicated automatically |
| Circulator S1 | cooling unit controls temperature of top plate of sample chamber temperature range = 14 - 28 °C temperature homogenity (microwell plate) = ± 2 °C available for air objectives only |
| Dispensing unit | offers on-axis access to specimen enables pipetting without disturbing environmental conditions allows sequential, semi-automatic pipetting of multi-positions |
| Insert plate for UV disinfection | incl. two UV bulbs, 1.0 W each emitting 254 nm fully automated disinfection process takes 23 min can be used on-demand or for preventive maintenance |











Count on Service in the True Sense of the Word

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Because the ZEISS microscope system is one of your most important tools, we make sure it is always ready to perform. What's more, we'll see to it that you are employing all the options that get the best from your microscope. You can choose from a range of service products, each delivered by highly qualified ZEISS specialists who will support you long beyond the purchase of your system. Our aim is to enable you to experience those special moments that inspire your work.

Repair. Maintain. Optimize.

Attain maximum uptime with your microscope. A ZEISS Protect Service Agreement lets you budget for operating costs, all the while reducing costly downtime and achieving the best results through the improved performance of your system. Choose from service agreements designed to give you a range of options and control levels. We'll work with you to select the service program that addresses your system needs and usage requirements, in line with your organization's standard practices.

Our service on-demand also brings you distinct advantages. ZEISS service staff will analyze issues at hand and resolve them – whether using remote maintenance software or working on site.

Enhance Your Microscope System.

Your ZEISS microscope system is designed for a variety of updates: open interfaces allow you to maintain a high technological level at all times. As a result you'll work more efficiently now, while extending the productive lifetime of your microscope as new update possibilities come on stream.







Profit from the optimized performance of your microscope system with services from ZEISS – now and for years to come.

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